Fungal Biology 124 (2020) 219–227

Contents lists available at ScienceDirect

Fungal Biology

journal homepage: www.elsevier.com/locate/funbio

Particle size distribution of the major *Alternaria alternata* allergen, Alt a 1, derived from airborne spores and subspore fragments

Łukasz Grewling [a, *], Paweł Bogawski [b], Agata Szymańska [a], Małgorzata Nowak [a], Łukasz Kostecki [a], Matt Smith [c]

[a] Laboratory of Aerobiology, Department of Systematic and Environmental Botany, Faculty of Biology, Adam Mickiewicz University, Uniwersytetu Poznańskiego 6, 61-614, Poznań, Poland
[b] Laboratory of Biological Spatial Information, Department of Systematic and Environmental Botany, Faculty of Biology, Adam Mickiewicz University, Uniwersytetu Poznańskiego 6, 61-614, Poznań, Poland
[c] School of Science and the Environment, University of Worcester, Henwick Grove, WR2 6AJ, Worcester, United Kingdom

ARTICLE INFO

Article history:
Received 10 October 2019
Received in revised form
7 February 2020
Accepted 9 February 2020
Available online 22 February 2020

Corresponding Editor: Nicholas P Money

Keywords:
Bioaerosols
Cascade impactor
ELISA
Fungal allergy
Hyphal fragments

ABSTRACT

Fungal fragments are abundant immunoreactive bioaerosols that may outnumber the concentrations of intact spores in the air. To investigate the importance of *Alternaria* fragments as sources of allergens compared to *Alternaria* spores, we determined the levels of *Alternaria* spores and Alt a 1 (the major allergen in *Alternaria alternata* spores) collected on filters within three fractions of particulate matter (PM) of different aerodynamic diameter: (1) $PM_{>10}$, (diameter>10 μm); (2) $PM_{2.5\text{-}10}$ (2.5–10μm); (3) $PM_{2.5}$ (0.12–2.5 μm). The airborne particles were collected using a three stage high-volume ChemVol cascade impactor during the *Alternaria* sporulation season in Poznań, Poland (30 d between 6 July and 22 September 2016). The quantification of Alt a 1 was performed using the enzyme-linked immunosorbent assay. High concentrations of Alt a 1 were recorded during warm and dry d characterized by high sunshine duration, lack of clouds and high dew point values. Atmospheric concentrations of *Alternaria* spores correlated significantly ($r = 0.930$, $p < 0.001$) with Alt a 1 levels. The highest Alt a 1 was recorded in $PM_{2.5\text{-}10}$ (66.8 % of total Alt a 1), while the lowest in $PM_{2.5}$ (<1.0 %). Significantly more Alt a 1 per spore (>30 %) was observed in $PM_{2.5\text{-}10}$ than in $PM_{>10}$. This Alt a 1 excess may be derived from sources other than spores, e.g. hyphal fragments. Overall, in outdoor air the major source of Alt a 1 are intact *Alternaria* spores, but the impact of other fungal fragments (hyphal parts, broken spores, conidiophores) cannot be neglected, as they may increase the total atmospheric Alt a 1 concentration.

1. Introduction

Fungal aerosols include generative and vegetative particles of different size and shapes, fragmented, whole or aggregated, that can be passively or actively released into the air (Afanou et al., 2014; Despres et al., 2012; Green et al., 2006). Airborne fungal particles are very common both in indoor and outdoor environment comprising a large proportion of total aerosol particle mass (Elbert et al., 2007; Frohlich-Nowoisky et al., 2009; Womiloju et al., 2003). The size of reproductive fungal propagules (spores, conidia) varies from approximately 1 μm to over 100 μm (Lacey and West, 2006). Airborne fragments of vegetative mycelium may be even smaller, reaching submicron dimensions (Green et al., 2006). Spores, conidiophores and hyphal fragments could be released simultaneously, but the releasing mechanism is different and depends on factors such as fungal species, weather conditions, mechanical disturbance (e.g. action of animals) as well as the texture, moisture and the vibration of the substrate (Afanou et al. 2014, 2015; Frankel et al., 2014; Górny et al., 2002; Green et al. 2005a, 2005b, 2006; Madsen et al., 2016; Magyar et al., 2016). Furthermore, morphological differences between intact spores and fungal fragments suggest that the atmospheric behaviour of these particles, e.g. deposition velocity, may vary substantially. For instance, measurement of fungal aerosols during aerosolisation experiments have shown that counts of fungal fragments do not always correlate well with spore concentrations (especially in low air velocity) (Górny et al., 2002).

* Corresponding author. Laboratory of Aerobiology, Department of Systematic and Environmental Botany, Faculty of Biology, Adam Mickiewicz University, Uniwersytetu Poznańskiego 6, 61-614, Poznań, Poland.
E-mail address: lukaszgrewling@gmail.com (Ł. Grewling).

https://doi.org/10.1016/j.funbio.2020.02.005

Fungal particles contain potentially harmful substances (mycotoxins and allergens) that may cause serious health problems (Rick et al., 2016). Development, persistence, and severity of allergic rhinitis and asthma have been associated with mold sensitivity, and it is estimated that around 6.5 million people worldwide have severe asthma with fungal sensitizations (Andersson et al., 2003; Denning et al., 2006; Knutsen et al., 2012). Allergenic proteins have been detected both in fungal spores (that are traditionally linked with allergic reactions) (Twaroch et al., 2015) and in hyphal fungal fragments (Green et al., 2005c; Levetin et al., 2009). The degree that these fragments function as sources of allergens and contribute to adverse health effects has not, however, been determined (Green et al., 2006). On the one hand, subspore fragments (fragments of hyphae and conidiophores, and broken spores) are abundant bioaerosols and due to their small size may stay airborne longer and penetrate into the lower regions of the respiratory tract more easily than larger intact spores (Cho et al., 2005; Lacey and West, 2006; Pady and Gregory, 1963; Pulimood et al., 2007). Also, as fragments have large surface area relative to their mass (in comparison to larger particles) they may show higher biological activity (Frankel et al., 2014). For instance, a positive association has been observed between asthma admission and the level of broken spores (but not hyphae) during thunderstorms (Pulimood et al., 2007). On the other hand, fungal allergens have predominantly been localized in the cell wall of mature spores, as shown for *Alternaria* sp. - one of the most clinically important fungal taxa (Burbach et al., 2009; Twaroch et al., 2012). Also, significant positive correlations have been found between atmospheric levels of the major *Alternaria alternata* allergen (Alt a 1) and both *Alternaria* spore concentrations (Agarwal et al., 1983; Da Silva et al., 2019) and allergy symptoms (Feo Brito et al., 2012) suggesting close dependencies between intact spores, allergens, and symptoms.

Intact spores, due to the action of unfavourable weather conditions or mechanical disturbances, may be additionally fragmented to enrich the subspore fraction of fungal aerosols (China et al., 2016; Pulimood et al., 2007). Also, many fungal spores are hygroscopic and can absorb water from the surrounding atmosphere. Eventually, due to osmotic shock, spores may burst in humid conditions releasing submicronic size fragments (China et al., 2016; Pasanen et al., 1991). Whether these fragments contain allergens remains unclear, but pollen-orientated studies revealed high amounts of pollen allergens in cytoplasmic content derived from fragmented pollen grains (Buters et al., 2015; Hoidn et al., 2005; Schäppi et al. 1997b, 1999; Taylor et al., 2002). The bursting of hyphal tips in water has also been documented (Bartnicki-Garcia and Lippman, 1972). This process resembles the abortive pollen germination in rainwater, when allergens are expelled from the tip of the pollen tube (Grote et al., 2003; Schäppi et al., 1997b). A recent study showed a positive correlation between Alt a 1 and *Alternaria* spores was only observed in buildings that had high relative humidity (Da Silva et al., 2019). In general, moisture is essential in fungal spore germination (Dagno et al., 2011; Hatzipapis et al., 2002; Vloutoglou et al., 1996), and it has been shown that *Alternaria* spores in extraction liquid release allergens in just 15 min (Sweeney et al., 1985). Germinating spores were also observed in the warm and moist environment of the nasal cavity, and this mechanism was postulated as an additional source of allergens (Green et al., 2003; Sercombe et al., 2006).

This study aims to determine whether subspore fragments of *Alternaria* can be a significant source of Alt a 1 in ambient air (in comparison to intact *Alternaria* spores) and to investigate the relationship between allergen content and environmental conditions, especially those related to moisture. This was achieved by examining the levels of Alt a 1 allergen released from airborne particles collected on filters within three fractions of particulate matter (PM) and relating these to various weather parameters.

2. Materials and methods

2.1. Fungal aerosol collection and quantification

Airborne *Alternaria* particles were collected using a high-volume (400 l/min) ChemVol cascade impactor (Butraco Inc., Son, Netherlands) (Buters et al., 2012; Demokritou et al., 2002). The ChemVol contains three impaction stages for collecting particles of different aerodynamic cut-off diameters i.e. (1) >10 μm ($PM_{>10}$); (2) 2.5–10 μm ($PM_{2.5\text{-}10}$); (3) 0.12–2.5 μm ($PM_{2.5}$). *Alternaria* spores are large with the aerodynamic diameters ranging from about 10 to 30 μm (McCartney et al., 1993). The ChemVol sampler therefore seemed a suitable collecting device for separation intact *Alternaria* spores from smaller fungal fragments (hyphal parts and dissected spores). *Alternaria* particles were collected in Poznań, during the main *Alternaria* sporulation season (30 d between 6 July and 22 September 2016, Tab. 1S). The highest daily *Alternaria* spore concentrations in Poznań are observed from the end of June to the middle of September, with the seasonal peak recorded usually in the beginning of August (Grewling et al., 2019; Kasprzyk et al., 2013). A previous study showed that the mean seasonal *Alternaria* spore concentration in Poznań was the highest among other cities in Central and Eastern Europe (Kasprzyk et al., 2015), which is related to the fact that the city is located in the agricultural region of Western Poland. Field crops are considered the main source (host plants) of various *Alternaria* species. In the studied area the predominant species are *A. alternata, Alternaria brassicicola,* and *Alternaria brassicae* as they infect oilseed rape fields that are abundant in Western Poland (Jajor et al., 2012; Baranowski et al., 2015). In addition, other crop pathogens are locally common, such as *Alternaria solani* (infecting potatoes), *Alternaria porri* (pathogen of onion), and *Alternaria dauci* that infects carrots (Gawińska-Urbanowicz and Kapsa, 2013; Ogórek et al., 2011). However, it should be stressed that spores of different *Alternaria* species are morphologically similar and so, when airborne samples are investigated, *Alternaria* spores cannot be identified to species level and are therefore grouped to genus level. The ChemVol was located at roof level (18 m a.g.l.) in the northern part of Poznań (52.46′N, 16.92′E) (Fig. S1). The sampling time was 24 h, from 12:00 to 12:00 of the next day, but is described as a “daily average” throughout. The collection substrates were polyurethane foam filters. Each filter (three filters per day) was cut into three equal pieces and extracted in the dark (4 h in 0.1 M ammonium bicarbonate buffer). After filter extraction the content was centrifuged (10 min at 1699 g). The supernatant was used for quantification of the major *A. alternata* allergen (Alt a 1), while sediment was used for *Alternaria* spore calculation. The quantification of Alt a 1 in each air fraction was performed using the enzyme-linked immunosorbent assay (ELISA) following the protocol described in Grewling et al. (2019). Due to a high degree of structural similarity between Alt a 1 homologues proteins (Amado and Barnes, 2016; Hong et al., 2005; Sáenz-de-Santamaría et al., 2006), fungal species closely related to *A. alternata* could also be detected by ELISA. Therefore, when describing the Alt a 1 concentration in the air the results are not limited only to *A. alternata*, but may also refer to other *Alternaria* species. Other fungal genera, e.g. *Urocladium* and *Stemphylium* that show close phylogenetic relationship to *Alternaria* (Gutierrez-Rodriguez et al., 2011), should not markedly affect the obtained results as concentrations of their spores in the air are often very low (Bednarz and Pawłowska, 2016; Ścevkova and Kovac, 2019). The daily mean Alt a 1 concentration was expressed as pg/m^3. The number of spores extracted from filters was calculated

using a method adopted from the estimation of pollen production (Bogawski et al., 2016). After centrifugation, the sediment with spores was diluted in 200 μl of distilled water. This was vortexed to obtain a homogenous solution and 25ul was transferred to a microscope slide and gently spread within 1.5 × 1.5 cm area. The spores were counted under a light microscope (magnification 200x) in three horizontal lines. Taking into account the microscope field of view and number of lines the total examined area was 49.5 mm^2, i.e. 22 % of total slide area (225 mm^2). This procedure was repeated three times and the number of spores was averaged. The obtained value was used to calculate the mean spore concentration in 1 μl of solution. We decided to not express the spore concentration in 1 m^3 of air due to the uncertainty in estimating the total number of spores collected on filters (this value cannot be precisely calculated using proposed method of spore extraction).

To validate the correctness of spore enumeration from filters, the results were compared with the mean "daily" level of spores (from 12:00 to 12:00 of the next day) obtained by routine methods used in aerobiology, i.e. volumetric Hirst spore trap (Hirst, 1952), located next to the ChemVol impactor. The Hirst (1952) spore trap is a impaction type sampler where air is sucked at a rate of 10 l/min through a 2 mm × 14 mm orifice. Behind the orifice the air flows over a rotating drum that moves past the inlet at 2 mm/h and is covered with an adhesive coated, transparent plastic tape. Airborne spores impact on the tape to give a time related sample. Following its removal from the trap, the tape is divided into segments corresponding to 24-h periods (48 mm in length). Each segment is mounted between a glass slide and cover slip, and the samples are examined by light microscopy (× 400 magnification). Spores were counted along two longitudinal transects following the method described in literature (Maya-Manzano et al., 2016). The correlation between both datasets was positive and statistically significant (Pearson correlation coefficient, $r = 0.930$, $p < 0.05$) ensuring that selected methods give comparable results. The usefulness of the ChemVol impactor for bioaerosols collection has also been validated during the EU funded HIALINE project (Buters et al., 2015).

2.2. Weather data collection

Weather data were retrieved from the official weather station of the National Institute of Meteorology and Water Management located at the Poznań Ławica airport (app. 5 km southwest from aerobiological station) (Fig. S1). The temporal resolution of weather data was adjusted to the Alt a 1 collection time (12:00–12:00). The following meteorological parameters were analysed: daily mean, minimum and maximum air temperature (°C), dew point (°C), vapour pressure deficit - VPD (kPa), relative humidity (%), rainfall (mm), sunshine duration (h), wind speed (m/s), cloudiness (unit 1/8), daily fraction of specific cloud types (%): *Cumulus humilis* (*Cu hum*)/*Cumulus fractus* (*Cu fra*), *Cumulus mediocris* (*Cu med*)/*Cumulus congestus* (*Cu con*), *Stratocumulus* (*Sc*), *Cumulonimbus capillatus* (*Cu cap*), and daily fraction of all types of clouds (%).

2.3. Statistical analysis

The concentrations of Alt a 1 and *Alternaria* spores collected in three air fractions were compared by the Kruskal–Wallis H test and Dunn's procedure for multiple pairwise comparison. *P*-values have been adjusted using Benjamini-Hochberg correction. Relationships between Alt a 1 levels and *Alternaria* spore concentrations in selected air fractions were checked by simple linear regression analysis. Daily mean concentration of Alt a 1 in every stage of ChemVol sampler has been correlated (by Pearson correlation coefficient) with meteorological parameters. Also, the ratio in the level of Alt a 1 (or spores) between three investigated air fractions has been correlated with meteorological parameters. Data that were right (positively) skewed have been transformed (log+1). The multivariate principal component analysis (PCA) was performed to select the major weather conditions affecting the daily Alt a 1 concentration. Days with Alt a 1 concentration were divided into three groups based on two cut points estimated via probability quantiles (33.3 % and 66.6 %), i.e.: "low" (daily Alt a 1 levels< 1.98 pg Alt a 1/m^3, n = 10), "medium" (1.98–6.71 pg Alt a 1/m^3, n = 10) and "high" concentration (>6.71 pg Alt a 1/m^3, n = 10). Before PCA analysis, the data were Box–Cox transformed, scaled and centred. All statistical analysis has been performed using computing environment R (R Core Team, 2018) and packages: FactoMineR (Lê et al., 2008), corrplot (Wei and Simko, 2017), caret (Kuhn, 2008), and factoextra (Kassambara and Mundt, 2017).

3. Results

3.1. Distribution of Alt a 1 in different fraction of particulate matter

The highest amount of Alt a 1 was detected in the 2.5–10 μm ($PM_{2.5-10}$) air fraction (Fig. 1) and was significantly higher ($p < 0.05$) than in two other air fractions (Fig. 2). The Alt a 1 concentration was extremely low (<1 % of total Alt a 1) in the $PM_{2.5}$ air fraction that contained the smallest particles (i.e. <2.5 μm). A similar pattern was observed in relation to *Alternaria* spores, as the highest spore level was observed in $PM_{2.5-10}$, while the lowest in $PM_{2.5}$. Airborne concentrations of Alt a 1 were significantly ($p < 0.001$) related to *Alternaria* spore levels collected in $PM_{>10}$, $PM_{2.5-10}$ and $PM_{2.5}$ ($R^2 = 0.801$, $R^2 = 0.819$, and $R^2 = 0.454$, respectively) (Fig. 3). The correlation between total Alt a 1 and *Alternaria* spores was positive and significant ($R^2 = 0.865$, $p < 0.001$). Significantly more Alt a 1 per spore (31.3 %) was observed in $PM_{2.5-10}$ than in $PM_{>10}$ ($p = 0.015$).

3.2. Impact of weather on Alt a 1 distribution

There were significant positive correlations ($p < 0.05$) between daily *Alternaria* spore levels collected in the Hirst type trap and daily mean, maximum and minimum temperature, VPD, dew point, and sunshine duration. Similar relationships were recorded between daily levels of Alt a 1 (collected by ChemVol sampler) and weather conditions (Fig. 4). For instance, the Alt a 1 in every air fraction correlated positively with daily maximum ($r > 0.387$, $p > 0.05$) and mean temperature ($r > 0.384$, $p > 0.05$). Furthermore, statistically significant positive correlations were recorded between Alt a 1 in the $PM_{2.5}$ fraction and fair weather *Cumulus humilis* clouds ($r = 0.584$, $p < 0.001$), sunshine duration ($r = 0.567$, $p = 0.001$), and VPD ($r = 0.505$, $p = 0.004$).

On the other hand, humidity ($r = -0.447$, $p = 0.013$), the occurrence of all types of clouds ($r = -0.552$, $p = 0.001$) and cloudiness ($r = -0.495$, $p = 0.005$) all had significant negative associations with levels of Alt a 1 in the $PM_{2.5}$ fraction. In addition, there was a significant negative correlation between Alt a 1 and wind speed ($r = -0.372$) in the larger fractions. There were no significant relationships between Alt a 1 concentrations and rainfall or *Cumulonimbus capillaris* clouds. Finally, no significant relationships have been observed between *Alternaria* spores and meteorological parameters related to "humid conditions", such as cloudiness, the occurrence of *Stratocumulus* and all types of clouds, rainfall and increased humidity.

Considering the impact of meteorological conditions on the ratio of Alt a 1 (or spores) recorded in different air fractions, only two significant correlations have been observed: (1) Alt a 1 in PM2.5 to Alt a 1 in >PM2.5 was negatively correlated with humidity ($r = -0.411$, $p = 0.024$); (2) Spores in PM2.5 to spores collected in

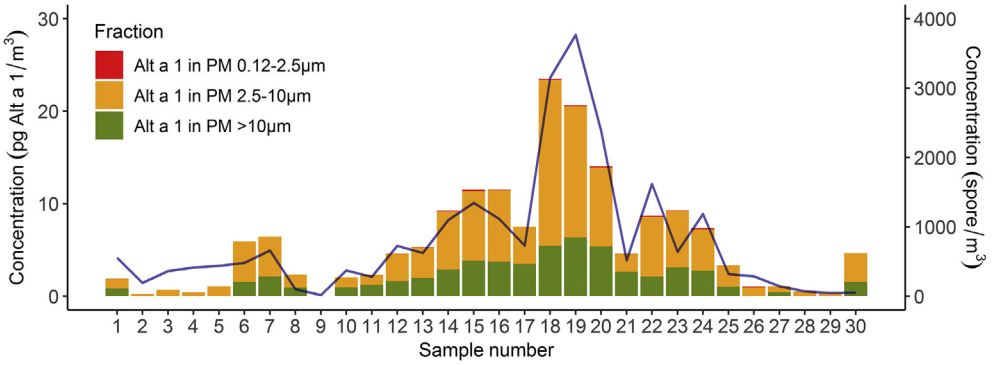

Fig. 1. Distribution of Alt a 1 in three investigated air fractions. *Alternaria* spores concentration (line curve) collected using Hirst type volumetric trap (samples description in Table S1).

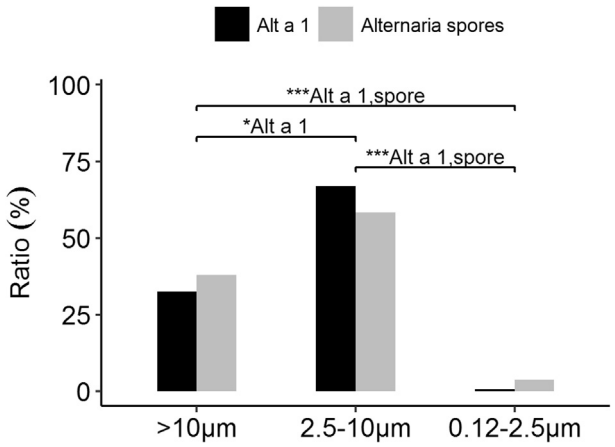

Fig. 2. Ratio (%) between the concentrations of Alt a 1 (pg/m^3) and *Alternaria* spores (spore/m^3) in selected air fractions to the total levels of Alt a 1 and total *Alternaria* spores. The statistically significant differences between spores and Alt a 1 levels in three air fractions are marked by asterisks, i.e. *p < 0.05, **p < 0.01, ***p < 0.001.

>PM2.5 correlated significantly with sunshine duration (r = −0.413, p = 0.023) (Fig. 5).

PCA supports results of correlation analysis, i.e. higher Alt a 1 concentrations were generally recorded during warm and dry d characterized by high sunshine duration (PC1), dew point and daily mean, maximum and minimum temperatures (PC2) (Fig. 6, Figs. S2 and S3). In addition, the occurrence of *Stratocumulus* clouds and all types of clouds (PC1) showed strong negative relationship with Alt a 1 concentrations. The first two principal components explained 65.7 % of variability in the dataset (Fig. S2).

4. Discussion

4.1. Sources of Alt a 1

Our study demonstrates that the daily levels of *Alternaria* spores correlated significantly with Alt a 1 (r = 0.930, p < 0.001), and we can therefore assume that the majority of atmospheric Alt a 1 was derived from intact *Alternaria* spores. Spores take part in the infection of plants and Alt a 1 is a protein involved in plant pathogenesis, i.e. interacts with plant defence proteins such as PR5 (Garrido-Arandia et al., 2016). In other words, spores need Alt a 1 to block plant defences and to favour fungal entry into the plant. In view of these findings, it is not surprising that Alt a 1 was located in the highest concentrations in the cell walls of old and germinating spores (Mitakakis et al., 2001; Twaroch et al., 2012) because the allergen is located exactly where it is most needed. The length of *Alternaria* spores vary from approximately 20 µm to as much as 200 µm (Simmons, 2007) and are many times larger than micronic sized fungal fragments. In addition, only part of the hyphal fragments are immunoreactive, as around 25 % of all hyphae expressed detectable allergens (Green et al., 2005c). Hundreds of fragments are therefore needed to exceed the allergen load of a single spore, and comparative studies showed that the differences between the levels of airborne hyphal fragments and spores are not in fact as high. According to Green et al. (2005c) fungal hyphae concentrations surpassed spores only by around 2–3 times. Higher differences (exceeding even 300-fold) have been observed in aerosolization experiments (Górny et al., 2002), but the mean difference between the number of hyphal fragments and spores was much lower (varying from 10 to 60-fold depending on air velocity). In addition, in a study conducted in two US cities, fungal fragments were present on 99 % of all d, although spores rather than hyphae

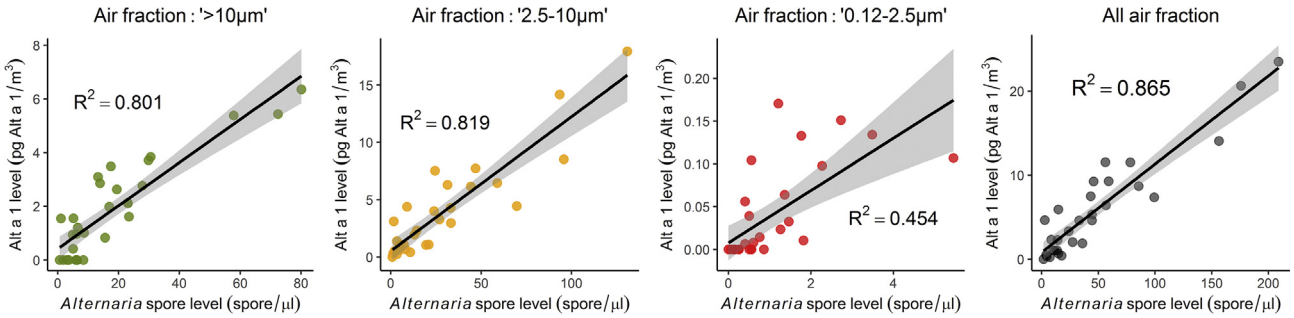

Fig. 3. Correlations between Alt a 1 concentrations and *Alternaria* spore levels in selected air fractions.

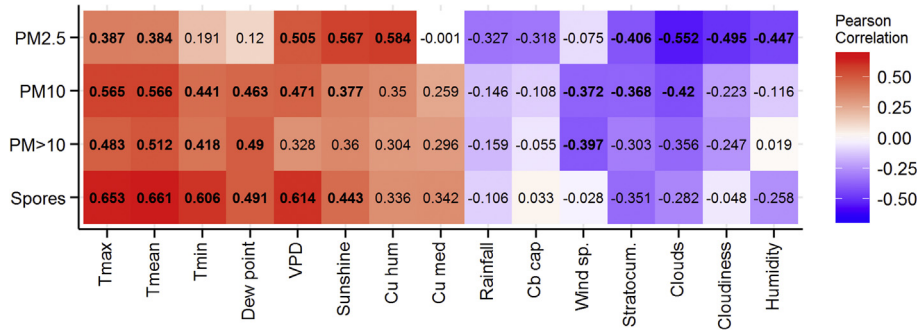

Fig. 4. Correlations matrix showing relationships between daily weather parameters and both *Alternaria* spores (collected by Hirst trap) and Alt a 1 in three investigated fractions of particulate matter (statistically significant correlations with $p < 0.05$ are in bold).

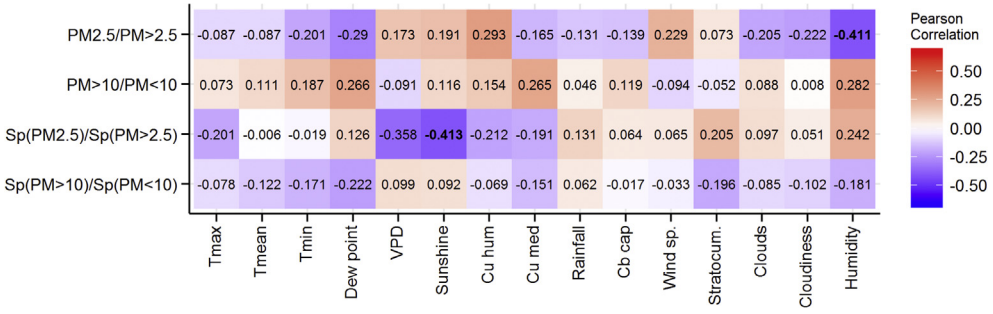

Fig. 5. Correlations matrix showing relationships between daily weather parameters and ratio in the level of Alt a 1 (or spores) between investigated air fraction of particulate matter (statistically significant correlations with $p < 0.05$ are in bold).

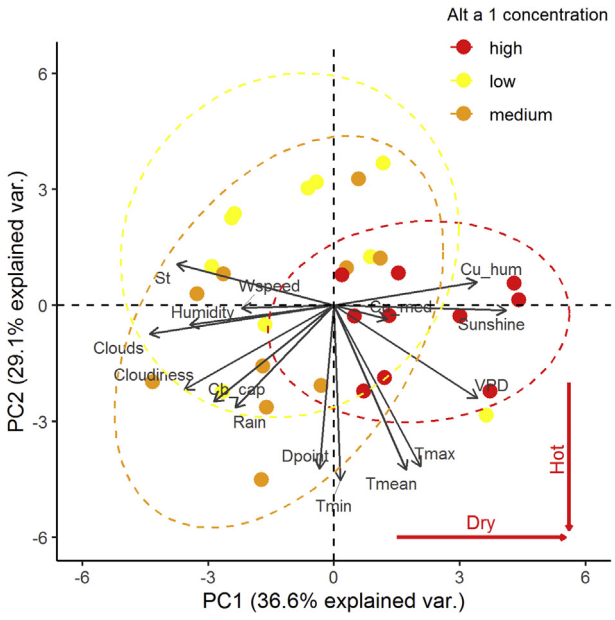

Fig. 6. Principal component analysis, using weather data collected during sampling period, for Alt a 1 concentration levels (ellipses represent the 90 % confidence interval of selected groups). Additional information of PCA analysis in Suppl. Materials, Fig. S2 & S3.

predominated in the air (Levetin et al., 2009). What is more, it should be stressed that in environmental samples, it is extremely difficult to morphologically distinguish the hyphae of different fungal species (immunostaining and DNA extraction techniques may be a solution) (Green et al., 2005b; Rittenour et al., 2012). In some of the studies mentioned previously (Green et al., 2005c; Levetin et al., 2009), only total hyphae fragments were counted (without species recognition). This approach, although valuable, does not allow for the direct comparison between spores and hyphae of particular fungal species, so the contribution of mycelial fragments could be overestimated.

The highest level of *Alternaria* spores (and Alt a 1) was observed in the $PM_{2.5\text{-}10}$ air fraction. This is surprising as, according to previous studies, the aerodynamic diameter of *Alternaria* spores exceeds 10 μm (McCartney et al., 1993; Yamamoto et al., 2014). Most of the spores should therefore be deposited in the $PM_{>10}$ air fraction, as it was presented in pollen-oriented studies (based on the same experimental setup) where around 90 % of pollen allergens (and therefore also pollen grains) were detected in $PM_{>10}$ fraction (Buters et al. 2012, 2015; Galán et al., 2013; Grewling et al., 2016). It is worth noting, however, that *Alternaria* colonies were also isolated from air samplers with particle diameter lower than 10 μm (Kim et al., 2010; Sayer et al., 1969). Furthermore, DNA barcoding analysis (Yamamoto et al., 2012) showed that the concentration of *Alternaria* DNA was also very high in the $PM_{2.5\text{-}10}$ fraction (although DNA might originate from both fungal spores and fragments). Presumably, the high number of *Alternaria* spores in the $PM_{2.5\text{-}10}$ fraction derived from their characteristic elongated club shape (in contrast to spherical pollen grains). McCartney et al. (1993) stressed that because of the shape of *Alternaria* spores, the mass is not uniformly distributed along their length, and thus it is difficult to predict their aerodynamic characteristics. In addition, the aerodynamic diameter of fungal spores cannot be accurately estimated solely based on the physical diameter but needs additional information, e.g. on the density of the spores and ambient air humidity (Reponen et al., 2001). For instance, it has been shown that high humidity may increase the diameter of *Cladosporium* spores by as much as 180 % (Pasanen et al., 1991).

Our study revealed that the proportion of Alt a 1 to spores in the $PM_{2.5-10}$ fraction was around 30 % higher than in $PM_{>10}$. Presumably, the 30 % excess of Alt a 1 in $PM_{2.5-10}$ derived from subspore hyphal fragments. However, it should be noted, that we did not quantify the level of *Alternaria* fungal fragments (based on their morphology) in air samples in this study (only Alt a 1 derived from fragments). When interpreting the peculiarities in allergen concentrations in the air, one should also remember about high variation in the allergenicity of fungal spores (Grewling et al., 2019; Mitakakis et al., 2001). Grewling et al. (2019) revealed differences of up to eightfold in day-to-day variations in *Alternaria* spore allergenicity that could be linked to varying species composition during the sporulation season. Spores of different *Alternaria* species vary in their aerodynamic properties (McCartney et al., 1993), and so species-specific variations in the aerodynamic behaviour of spores (with different Alt a 1 content) may also affect the amount of Alt a 1 recorded in different fractions of particulate matter.

4.2. Impact of weather on Alt a 1 concentration

The results of PCA and correlation analysis showed that the highest Alt a 1 levels were recorded during sunny, warm and dry d, when weather conditions favoured the upward movement of air currents (high temperature, sunshine duration, dew point and VPD, and presence of *Cumulus* clouds). These conditions are known to positively affect the daily concentrations of *Alternaria* spores (Grinn-Gofroń and Bosiacka, 2015; Hjelmroos, 1993; Stennett and Beggs, 2004; Troutt and Levetin, 2001). It is striking to note that the strength and direction of correlations between meteorological factors and both Alt a 1 (especially in $PM_{>10}$ and $PM_{2.5-10}$ fractions) and *Alternaria* spore concentrations were generally very similar (see Fig. 4), which supports the idea that the main sources of Alt a 1 are *Alternaria* spores.

In contrast to findings from pollen-oriented studies where humid conditions increased the fraction of allergens related to small fragments (Buters et al., 2015; Schäppi et al., 1997a), our experiment showed an opposite situation. The ratio of Alt a 1 in $PM_{2.5}$ to Alt a 1 in larger fractions increased with decreasing relative humidity. This result concurs with previous findings (Madsen, 2012) showing that the fraction of the fungal particles being of respirable size was the highest for particles aerosolized at low relative humidity. We suspect that the different behaviour of pollen and fungal spores arises from differences in the wall structure and the role of water in pollen/fungal spore germination. The release of subpollen allergens through pollen wall bursting (due to high humidity) was mainly described for pollen grains characterized by a delicate and thin pollen wall like grasses (Poaceae) (Schäppi et al., 1999; Taylor et al., 2002; Buters et al., 2015). It has not, however, been documented in pollen with thicker walls like mugwort (*Artemisia* sp.). In this species the concentration of subspore particle does not show positive relationship with air humidity (Grewling et al., 2020). Water has an adverse effect on pollen longevity and viability and so plants developed certain protective mechanisms (e.g. specific floral morphology, production of germination inhibitors) to prevent pollen from water damaging or undesirable germination (outside the stigma) (Eisikowitch and Woodell, 1974; Mao and Huang, 2009). In contrast, water is essential for fungal spore germination, and spores of some *Alternaria* species only germinate at 100 % relative humidity (Dickinson and Bottomley, 1980; Hatzipapis et al., 2002). Fungal spores are therefore adapted to moisture conditions and likely more resistant to rupturing by osmotic shock than thin-wall pollen grains. The recent study by Lawler et al. (2020) documented the occurrence of fungal nanoparticles in the air, which peaked around 1.5 d after the rainfall. Similar behaviour was also observed in the Amazon, where daily increase in fungal particles was related to high nighttime relative humidity (China et al., 2016). This suggests that post-rain processes related to fungal spore germination may play a role in the release of nanoparticles (Lawler et al., 2020). Considering the fragmentation of *Alternaria* spores, the mechanical damage to spores (e.g. observed during grass moving or harvesting (Pulimood et al., 2007)) seems to be more important than osmotic rupture.

In addition, our study showed that fine fungal fragments were more likely, than larger particles, to become airborne when *Cumulus humilis* clouds were observed. This type of cloud indicates unstable atmospheric conditions below the clouds, especially during their formation. Such turbulent conditions could occur during intense sunshine duration preceding *Cumulus humilis* formation and/or in the presence of *C. hum* clouds (Stull, 1985). Indeed, *C. hum* and sunshine duration are highly positively correlated with the amount of Alt a 1 in smallest air fraction (see Fig. 4). Consequently, higher numbers of small, immunoreactive fungal particles may occur during weak or moderate convection (e.g. warm air ascending with velocity 2–5 m s^{-1}). Larger fungal fragments (>2.5 μm) therefore seem to be more loosely connected with weak convection, probably because they require stronger air movements to overcome gravity and drag.

Deposition velocity of airborne particles is the lowest (<0.03 cm s^{-1}) for particles of aerodynamic diameter between 0.1 and 1.0 μm. For larger particles (>5.0 μm) deposition velocity strongly increases (>1.0 cm s^{-1}) and sedimentation becomes a predominant atmospheric process of particle removal (Nicholson, 1995). According to Woo et al. (2018) *Alternaria* spores with aerodynamic diameter of 10 μm had a deposition velocity of 0.63 cm s^{-1}. Fine hyphal fragments (0.12–2.5 μm) might hypothetically gain in importance in indoor environments (Górny et al., 2002) or during specific weather conditions, e.g. thunderstorm events when many particles are uplifted, mixed and damaged (D'Amato et al., 2017; Pulimood et al., 2007). In our study, we investigated the effect of "storm-like" conditions, e.g. presence of *Cumulonimbus* clouds on Alt a 1 concentrations, but no significant relationships have been observed. Also, episodes of rain were uncommon during the study period and so we could not test the hypothesis linking occurrence of light rainfall (<1 mm) with increased level of allergens (Schäppi et al. 1997a, 1997b).

4.3. Alt a 1 in finest air fraction (PM 0.12–2.5 μm)

Previous studies have shown that the concentration of the smallest fungal fragments (~1 μm), in comparison to spores, can be relatively high in the air (Reponen et al., 2007; Adhikari et al., 2009; Lee and Liao, 2014). In these studies, the detection and enumeration of fungal fragments was mainly based on the concentration of (1 → 3)-ß-D-glucan, i.e. polysaccharide abundant in fungal cell walls (Rylander, 1999). Such methods do not, however, allow fragments belonging to specific fungal taxa to be identified, so all fungal fragments were grouped and counted together. When a molecular technique was applied, no sign of *Alternaria* DNA was found in the $PM_{2.5}$ fraction (Yamamoto et al., 2012). In the current study, we used a family-specific allergen that occurs in *A. alternata* and other members of Pleosporaceae family (as Alt a 1 homologues) (Hong et al., 2005; Sáenz-de-Santamaría et al., 2006). Based on this detection method, the total level of Alt a 1 in 0.12–2.5 μm air fraction was extremely low (1 % of total Alt a 1). This result concurs with pollen allergen studies where no allergens was found in the $PM_{2.5}$ fraction (Buters et al. 2010, 2012). Green et al. (2005b) postulated that the amount of allergens released from a hyphal fragment might be a function of the critical fragment size, which is the minimum size at which a fungal fragment remains viable. In the study conducted by Górny et al. (2002) it was shown that

immunoreactive fungal fragments of *Aspergillus, Penicillium* and *Cladosporium* might be as small as 0.3 μm. The critical sizes for *Alternaria* species have not, however, been established. In addition, Buters et al. (2010) showed that airborne pollen allergens of micronic size are easily absorbed by diesel soot particles. It was postulated that this phenomenon could be responsible for the lack of pollen allergens in micron-sized air fraction (de Weger et al., 2013). These studies and our results suggest that the vast majority of immunoreactive airborne *Alternaria* particles belongs to spores and larger fungal fragments.

5. Conclusions

Our study showed that the main source of airborne Alt a 1 in the outdoor environment are intact *Alternaria* spores (app. 80 %), which are deposited in both the $PM_{>10}$ and $PM_{2.5-10}$ air fractions. The possible contribution of other fungal particles is the most visible in $PM_{2.5-10}$, where fungal fragments may be responsible for more than 30 % of total Alt a 1. The amount of allergen related to the finest fungal fragments ($PM_{2.5}$) is very low, almost negligible from clinical and epidemiological point of view. This is important news, as the quantification of *Alternaria* spores in the air (without mycelial fragments) is currently a routine practice in many aerobiological laboratories. Our results suggest that such information could be used as a relevant approximation of exposure to airborne Alt a 1 (based on very strong correlation between spores and Alt a 1). Nevertheless, high variations in allergen content between individual spores should also be considered to fully evaluate the exposure level to *Alternaria* allergens.

Declaration of Competing Interest

The authors report no conflicts of interest. The authors alone are responsible for the content and the writing of the paper.

Acknowledgements

The study was supported by the Polish National Science Centre grant no. 2013/09/D/NZ7/00358. Authors would like to thank Karolina Mizeraczyk and Julia Gwiazdowska for their help with samples preparations.

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.funbio.2020.02.005.

References

Adhikari, A., Jung, J., Reponen, T., Lewis, J.S., DeGrasse, E.C., Grimsley, L.F., Chew, G.L., Grinshpun, S.A., 2009. Aerosolization of fungi, (1→3)-ß-D-glucan, and endotoxin from flood-affected materials collected in New Orleans homes. Environ. Res. 109 (3), 215–224.

Afanou, K.A., Straumfors, A., Skogstad, A., Nilsen, T., Synnes, O., Skaar, I., Hjeljord, L., Tronsmo, A., Green, B.J., Eduard, W., 2014. Submicronic fungal bioaerosols: high-resolution microscopic characterization and quantification. Appl. Environ. Microbiol. 80 (22), 7122–7130.

Afanou, K.A., Straumfors, A., Skogstad, A., Skaar, I., Hjeljord, L., Skare, O., Green, B.J., Tronsmo, A., Eduard, W., 2015. Profile and morphology of fungal aerosols characterized by field emission scanning electron microscopy (FESEM). Aerosol. Sci. Technol. 49 (6), 423–435.

Agarwal, M.K., Swanson, M.C., Reed, C.E., Yunginger, J.W., 1983. Immunochemical quantitation of airborne short ragweed, Alternaria, antigen E, and Alt-I allergens: a two-year prospective study. J. Allergy Clin. Immunol. 72 (1), 40–45.

Amado, M., Barnes, C., 2016. Allergenic microfungi and human health: a review on exposure, sensitization, and sequencing allergenic proteins. In: Biology of Microfungi. Springer, pp. 429–449.

Andersson, M., Downs, S., Mitakakis, T.Z., Leuppi, J., Marks, G., 2003. Natural exposure to Alternaria spores induces allergic rhinitis symptoms in sensitized children. Pediatr. Allergy Immunol. 14, 100–105.

Baranowski, P., Jedryczka, M., Mazurek, W., Babula-Skowrońska, D., Siedliska, A., Kaczmarek, J., 2015. Hyperspectral and thermal imaging of oilseed rape (Brassica napus) response to fungal species of the genus Alternaria. PloS One 31 (10), e0122913, 3.

Bartnicki-Garcia, S., Lippman, E., 1972. The bursting tendency of hyphal tips of fungi: presumptive evidence for a delicate balance between wall synthesis and wall lysis in apical growth. J. Gen. Microbiol. 73, 487–500.

Bednarz, A., Pawłowska, S., 2016. A fugal calendar for the atmosphere of Szczecin, Poland. Acta Agrobot. 69 (3), 1–9.

Bogawski, P., Grewling, Ł., Frątczak, A., 2016. Flowering phenology and potential pollen emission of three Artemisia species in relation to airborne pollen data in Poznań (Western Poland). Aerobiologia 32, 265–276.

Burbach, G.J., Heinzerling, L.M., Edenharter, G., Bachert, C., Bindslev-Jenses, C., Bonini, S., Bousquet, J., Bousquet-Rouanet, L., Bousquet, P.J., Bresciani, M., Bruno, A., Canonica, G.W., Darsow, U., Demoly, P., Durham, S., Fokkens, W.J., Giavi, S., Gjomarkaj, M., Gramiccioni, C., Haahtela, T., Kowalski, M.L., Magyar, P., Murakozi, G., Orosz, M., Papadopoulos, N.G., Rohnelt, C., Stingl, G., Todo-Bom, A., Von Mutius, E., Wiesner, A., Wohrl, S., Zuberbier, T., 2009. GA2LEN skin test study II: clinical relevance of inhalant allergen sensitizations in Europe. Allergy 64, 1507–1515.

Buters, J., Prank, M., Sofiev, M., Pusch, G., Albertini, R., Annesi-Maesano, I., Antunes, C., Behrendt, H., Berger, U., Bandao, R., Celenk, S., Galan Soldevilla, C., Grewling, Ł., Jackowiak, B., Kennedy, R., Rantio Lehtimäki, A., Resse, G., Sauliene, I., Smith, M., Thibaudon, M., Weber, B., Cecchi, L., 2015. Variation of the group 5 grass pollen allergen content of airborne pollen in relation to geographic location and time in season. J. Allergy Clin. Immunol. 136 (1), 87–95 e6.

Buters, J., Thibaudon, M., Smith, M., Kennedy, R., Rantio Lehtimäki, A., Albertini, R., Reese, G., Weber, B., Galán, C., Brandao, R., Antunes, C., Jäger, S., Berger, U., Celenk, S., Grewling, Ł., Jackowiak, B., Sauliene, I., Weichenmeier, I., Pusch, G., Sarioglu, H., Ueffing, M., Behrendt, H., Prank, M., Sofiev, M., Cecchi, L., 2012. Release of Bet v 1 from birch pollen from 5 European countries. Results from the HIALINE study. Atmos. Environ. 55, 496–505.

Buters, J.T., Weichenmeier, M.I., Ochs, S., Pusch, G., Kreyling, W., Boere, A.J.F., Schober, W., Behrendt, H., 2010. The allergen Bet v 1 in fractions of ambient air deviates from birch pollen counts. Allergy 65, 850–858.

China, S., Wang, B., Weis, J., Rizzo, L., Brito, J., Cirino, G.G., Kovarik, L., Artaxo, P., Gilles, M.K., Laskin, A., 2016. Rupturing of biological spores as a source of secondary particles in Amazonia. Environ. Sci. Technol. 50 (22), 12179–12186.

Cho, S.-H., Seo, S.-C., Schmechel, D., Grinshpun, S.A., Reponen, T., 2005. Aerodynamic characteristics and respiratory deposition of fungal fragments. Atmos. Environ. 39, 5454–5465.

D'Amato, G., Annesi-Maesano, I., Molino, A., Vitale, C., D'Amato, M., 2017. Thunderstorm-related asthma attacks. J. Allergy Clin. Immunol. 139 (6), 1786–1787.

Dagno, K., Lahlali, R., Diourte, M., Jijakli, M.H., 2011. Effect of temperature and water activity on spore germination and mycelial growth of three fungal biocontrol agents against water hyacinth (*Eichhornia crassipes*). J. Appl. Microbiol. 110 (2), 521–528.

Da Silva, H.A., Divjan, A., Dannemiller, K.C., Acosta, L.M., Miller, R.L., Green, B.J., Perzanowski, M., 2019. Correlations between Alt a 1 concentrations, *Alternaria alternata* and Pleasporaceae measured by molecular methods in New York City house dust. J. Allergy Clin. Immunol. 143 (2). Supplement, Page AB122.

de Weger, L.A., Bergmann, K.C., Rantio Lehtimäki, A., Dahl, A., Buters, J., Dechamp, C., Belmonte, J., Thibaudon, M., Cecchi, L., Besancenot, J.-P., Galan, C., Waisel, Y., 2013. Impact of pollen. In: Sofiev, M., Bergmann, K.C. (Eds.), Allergenic Pollen. A Review of the Production, Release. *Distribution and Health Impacts* Springer, Dordrecht.

Demokritou, P., Kavouras, I.G., Ferguson, S.T., Koutrakis, P., 2002. Development of a high volume cascade impactor for toxicological and chemical characterization studies. Aerosol. Sci. Technol. 36, 925–933.

Denning, D.W., O'Driscoll, R., Hogaboam, C.M., Bowyer, P., Niven, R.M., 2006. The link between fungi and severe asthma: a summary of the evidence. Eur. Respir. J. 27, 615–626.

Despres, V.R., Huffman, J.A., Burrows, S.M., Hoose, C., Safatov, A.S., Buryak, G., Frohlich-Nowoisky, J., Elbert, W., Andreae, M.O., Poschl, U., Jaenicke, R., 2012. Primary biological aerosol particles in the atmosphere: a review. Tellus B 64 (15598).

Dickinson, C.H., Bottomley, D., 1980. Germination and growth of alternaria and cladosporium in relation to their activity in the phylloplane. Trans. Br. Mycol. Soc. 74, 309–319.

Eisikowitch, D., Woodell, S.R.J., 1974. The effect of water on pollen germination in two species of primula. Evolution 28, 692–694.

Elbert, W., Taylor, P.E., Andreae, M.O., Poschl, U., 2007. Contribution of fungi to primary biogenic aerosols in the atmosphere: wet and dry discharged spores, carbohydrates, and inorganic ions. Atmos. Chem. Phys. 7 (17), 4569–4588.

Feo Brito, F., Alonso, A.M., Carnes, J., Martin-Martin, R., Fernandez-Caldas, E., Galindo, P.A., Alfaya, T., Amo-Salas, M., 2012. Correlation between Alt a 1 levels and clinical symptoms in Alternaria alternata-monosensitized patients. J. Invest. Allergol. Clin. Immunol. 22 (3), 154–159.

Frankel, M., Hansen, E.W., Madsen, A.M., 2014. Effect of relative humidity on the aerosolization and total inflammatory potential of fungal particles from dust-inoculated gypsum boards. Indoor Air 24 (1), 16–28.

Frohlich-Nowoisky, J., Pickersgill, D.A., Despres, V.R., Poschl, U., 2009. High diversity of fungi in air particulate matter. Proc. Natl. Acad. Sci. U.S.A. 106 (31), 12814–12819.

Galán, C., Antunes, C., Brandao, R., Torres, C., Garcia-Mozo, H., Caeiro, E., Ferro, R., Prank, M., Sofiev, M., Albertini, R., Berger, U., Cecchi, L., Celenk, S., Grewling, Ł., Jackowiak, B., Jäger, S., Kennedy, R., Rantio-Lehtimäki, A., Reese, G., Sauliene, I., Smith, M., Thibaudon, M., Weber, B., Weichenmeier, I., Pusch, G., Buters, J.T.M., 2013. Airborne olive pollen counts are not representative of exposure to the major olive allergen Ole e 1. Allergy 68, 809–812.
Garrido-Arandia, M., Silva-Navas, J., Ramirez-Castillejo, C., Cubells-Baeza, N., Gomez-Casado, C., Barber, D., Pozo, J.C., Melendi, P.G., Pacios, L.F., Diaz-Perales, A., 2016. Characterisation of a flavonoid ligand of the fungal protein Alt a 1. Sci. Rep. 6, 3346.
Gawińska-Urbanowicz, H., Kapsa, J., 2013. Monitoring results of population *Alternaria* genus in potato crops. Prog. Plant Protect. 53 (3), 527–532.
Górny, R.L., Reponen, T., Willeke, K., Schmechel, D., Robine, E., Boissier, M., Grinshpun, S.A., 2002. Fungal fragments as indoor air biocontaminants. Appl. Environ. Microbiol. 68 (7), 3522–3531.
Green, B.J., Mitakakis, T.Z., Tovey, E.R., 2003. Allergen detection form 11 fungal species before and after germination. J. Allergy Clin. Immunol. 111 (2), 285–289.
Green, B.J., Schmechel, D., Sercombe, J.K., Tovey, E.R., 2005a. Enumeration and detection of aerosolized Aspergillus fumigatus and Penicillium chrysogenum conidia and hyphae using a novel double immunostaining technique. J. Immunol. Methods 307, 127–134.
Green, B.J., Schmechel, D., Tovey, E.R., 2005b. Detection of aerosolized *Alternaria alternata* conidia, hyphae, and fragments by using a novel double-immunostaining technique. Clin. Diagn. Lab. Immunol. 12 (9), 1114–1116.
Green, B.J., Sercombe, J.K., Tovey, E.R., 2005c. Fungal fragments and undocumented conidia function as new aeroallergen sources. J. Allergy Clin. Immunol. 115, 1043–1048.
Green, B.J., Tovey, E.R., Sercombe, J.K., Blachere, F.M., Beezhold, D.H., Schmechel, D., 2006. Airborne fungal fragments and allergenicity. Med. Mycol. 44 (Suppl. 1), 245–255.
Grewling, Ł., Bogawski, P., Jenerowicz, D., Czarnecka-Operacz, M., Sikoparija, B., Skjøth, C.A., Smith, M., 2016. Mesoscale atmospheric transport of ragweed pollen allergens from infected to uninfected areas. Int. J. Biometeorol. 60 (10), 1493–1500.
Grewling, Ł., Nowak, M., Szymańska, A., Kostecki, Ł., Bogawski, P., 2019. Temporal variability in the allergenicity of airborne *Alternaria* spores. Med. Mycol. 4, 403–411.
Grewling, Ł., Bogawski, P., Kostecki, Ł., Nowak, M., Szymańska, A., Frątczak, A., 2020. Atmospheric exposure to the major *Artemisia* pollen allergen (Art v 1): seasonality, impact of weather, and clinical implications. Sci. Total Environ. 713, 136611.
Grinn-Gofroń, A., Bosiacka, B., 2015. Effects of meteorological factors on the composition of selected fungal spores in the air. Aerobiologia 31 (1), 63–72.
Grote, M., Valenta, R., Reichelt, R., 2003. Abortive pollen germination: a mechanism of allergen release in birch, alder, and hazel revealed by immunogold electron microscopy. J. Allergy Clin. Immunol. 111 (5), 1017–1023.
Gutierrez-Rodriguez, A., Postigo, I., Guisantes, J.A., Sunen, E., Martinez, J., 2011. Identification of allergens homologous to Alt a 1 from Stemphylium botryosum and Ulocladium botrytis. Med. Mycol. 49, 892–896.
Hatzipapis, P., Kalosak, K., Dara, A., Christias, C., 2002. Spore germination and appressorium formation in the entomopathogenic *Alternaria alternata*. Mycol. Res. 106 (11), 1349–1359.
Hirst, J.M., 1952. An automatic volumetric spore trap. Ann. Appl. Biol. 39, 257–265.
Hjelmroos, M., 1993. Relationship between airborne fungal spore presence and weather variables. Grana 32, 40–47.
Hoidn, C., Puchner, E., Pertl, H., Holztrattner, E., Obermeyer, G., 2005. Nondiffusional release of allergens from pollen grains of *Artemisia vulgaris* and *Lilium longiflorum* depends mainly on the type of the allergen. Int. Arch. Allergy Immunol. 137, 27–36.
Hong, S.G., Cramer, R.A., Lawrence, C.B., Pryor, B.M., 2005. Alt a 1 allergen homologs from Alternaria and related taxa: analysis of phylogenetic content and secondary structure. Fungal Genet. Biol. 42 (2), 119–129.
Jajor, E., Kozłowska, M., Wójtowicz, M., 2012. Prevalence of fungi of the genus *Alternaria* on rape siliques and seeds depending on weather conditions. Progress in Pland Protection 52 (4), 1011–1015.
Kasprzyk, I., Sulborska, A., Nowak, M., Szymańska, A., Kaczmarek, J., Haratym, W., Weryszko-Chmielewska, E., Jędryczka, M., 2013. Fluctuation range of the concentration of airborne *Alternaria* conidiospores sampled at different geographical locations in Poland (2010-2011). Acta Agrobot. 66 (1), 65–76.
Kasprzyk, I., Rodinkova, V., Šauliene, I., Ritenberga, O., Grinn-Gofroń, A., Nowak, M., Sulborska, A., Kaczmarek, J., Weryszko-Chmielewska, E., Bilous, E., Jędryczka, M., 2015. Air pollution by allergenic spores of the genus *Alternaria* in the air of central and eastern Europe. Environ. Sci. Pollut. Control Ser. 22, 9260–9274.
Kassambara, A., Mundt, F., 2017. Factoextra: Extract and Visualize the Results of Multivariate Data Analyses, 1.0.4. In: R Package. CRAN. Retrieved from. http://www.sthda.com/english/rpkgs/factoextra.
Kim, K.Y., Kim, Y.S., Kim, D., 2010. Distribution characteristics of airborne bacteria and fungi in the general hospitals of Korea. Ind. Health 48, 236–243.
Knutsen, A.P., Bush, R.K., Demain, J.G., Denning, D.W., Dixit, A., Fairs, A., Greenberger, P.A., Kariuki, B., Kita, H., Kurup, V.P., Moss, R.B., Niven, R.M., Pashley, C.H., Slavin, R.G., Vijay, H.M., Wardlaw, A.J., 2012. Fungi and allergic lower respiratory tract diseases. J. Allergy Clin. Immunol. 129 (2), 280–291.
Kuhn, M., 2008. Building predictive models in R using the caret package. J. Stat. Software 28 (5), 1–26.
Lacey, M.E., West, J.S., 2006. The Air Spora. A Manual for Catching and Identifying Airborne Biological Particles. Springer, Dordrecht, The Netherlands.
Lawler, M.J., Draper, D.C., Smith, J.N., 2020. Atmospheric fungal nanoparticle bursts. Science Advances 6, eaax9051.
Lê, S., Josse, J., Husson, F., 2008. FactoMineR: an R package for multivariate analysis. J. Stat. Software 25 (1), 1–18.
Lee, S.-A., Liao, C.-H., 2014. Size-selective assessment of agricultural workers' personal exposure to airborne fungi and fungal fragments. Sci. Total Environ. 466–467, 725–732.
Levetin, E., Owens, C., Weaver, H., Davis, W., 2009. Airborne fungal fragments: are we overlooking an important source of aeroallergens? J. Allergy Clin. Immunol. 123 (2), S231.
Madsen, A.M., 2012. Effects of airflow and changing humidity on the aerosolization of respirable fungal fragments and conidia of *Botrytis cinerea*. Appl. Environ. Microbiol. 78 (11), 3999–4007.
Madsen, A.M., Larsen, S.T., Koponen, I.K., Kling, K.I., Barooni, A., Karottki, D.G., Tendal, K., Wolkoff, P., 2016. Generation and characterization of indoor fungal aerosols for inhalation studies. Appl. Environ. Microbiol. 82 (8), 2479–2493.
Magyar, D., Vass, M., Li, D.W., 2016. Dispersal strategies of microfungi. In: Li, D.W. (Ed.), *Biology of Microfungi*. Fungal Biology. Springer, Cham.
Mao, Y.-Y., Huang, S.-Q., 2009. Pollen resistance to water in 80 angiosperm species: flower structures protect rain-susceptible pollen. New Phytol. 183, 892–899.
Maya-Manzano, J.M., Munoz-Trivino, M., Fernandez-Rodriguez, S., Silva-Palacios, I., Gonzalo-Garijo, A., Tormo-Molina, R., 2016. Airborne *Alternaria* conidia in Mediterranean rural environments in SW of Iberian Peninsula and weather parameters that influence their seasonality in relation to climate change. Aerobiologia 32 (1), 95–108.
McCartney, H.A., Schmechel, D., Lacey, M.E., 1993. Aerodynamic diameter of conidia of *Alternaria* species. Plant Pathol. 42 (2), 280–286.
Mitakakis, T.Z., Barnes, C., Tovey, E.R., 2001. Spore germination increases allergen release from Alternaria. J. Allergy Clin. Immunol. 107, 388–390.
Nicholson, K.W., 1995. Physical aspects of bioaerosol sampling and deposition. In: Cox, ChS., Wathes, ChM. (Eds.), Bioaerosols Handbook. Lewis Publishers, Boca Raton.
Ogórek, E., Pląskowska, E., Kalinowska, K., 2011. Characteristics and taxonomy of *Alternaria* fungi. Mikol. Lek. 18 (3), 150–155.
Pady, S.M., Gregory, P.H., 1963. Numbers and viability of airborne hyphal fragments in England. Trans. Br. Mycol. Soc. 46 (4), 609–613.
Pasanen, A.-L., Pasanen, P., Jantunen, M.J., Kalliokoski, P., 1991. Significance of air humidity and air velocity for fungal spore release into the air. Atmos. Environ. 25A (2), 459–462.
Pulimood, T.B., Corden, J., Bryden, C., Sharples, L., Nasser, S., 2007. Epidemic asthma and the role of the fungal mold *Alternaria alternata*. J. Allergy Clin. Immunol. 120 (3), 610–617.
R Core Team, 2018. R: *A Language and Environment for Statistical Computing* (Vienna, Austria).
Reponen, T., Grinshpun, S.A., Conwell, K.L., Wiest, J., Anderson, M., 2001. Aerodynamic versus physical size of spores: measurement and implication for respiratory deposition. Grana 40, 119–125.
Reponen, T., Seo, S.-C., Grimsley, F., Lee, T., Crawford, C., Grinshpun, S.A., 2007. Fungal fragments in moldy houses: a field study in homes in New Orleans and Southern Ohio. Atmos. Environ. 41 (37), 8140–8149.
Rick, E.M., Woolnough, K., Pashley, C.H., Wardlaw, A.J., 2016. Allergic fungal airway disease. J. Invest. Allergol. Clin. Immunol. 26 (6), 344–354.
Rittenour, W.R., Park, J.-H., Cox-Ganser, J.M., Beezhold, D.H., Green, B.J., 2012. Comparison of DNA extraction methodologies used for assessing fungal diversity via ITS sequencing. J. Environ. Monit. 14 (3), 766–774.
Rylander, R., 1999. Indoor air-related effects and airborne (1→3)-ß-D-glucan. Environ. Health Perspect. 107 (Suppl. 3), 501–503.
Sáenz-de-Santamaría, M., Postigo, I., Gutierrez-Rodriguez, A., Cardona, G., Guisantes, J.A., Asturias, J., Martinez, J., 2006. The major allergen of *Alternaria* alternata (Alt a 1) is expressed in other members of the Pleosporaceae family. Mycoses 49 (2), 91–95.
Sayer, W.J., Shean, D.B., Ghosseiri, J., 1969. Estimation of airborne fungal flora by the Andersen sampler versus the gravity settling culture plate. J. Allergy 44 (4), 214–227.
Ščevkova, A., Kovac, J., 2019. First fungal spore calendar for the atmosphere of Bratislava, Slovakia. Aerobiologia 35, 343–356.
Schäppi, G.F., Suphioglu, C., Taylor, P.E., Knox, R.B., 1997a. Concentrations of the major birch tree allergen Bet v 1 in pollen and respirable fine particles in the atmosphere. J. Allergy Clin. Immunol. 100 (5), 656–661.
Schäppi, G.F., Taylor, P.E., Staff, I.A., Suphioglu, C., Knox, R.B., 1997b. Source of Bet v 1 loaded inhalable particles from birch releaved. Sex. Plant Reprod. 10, 315–323.
Schäppi, G.F., Taylor, P.E., Pain, M.C.F., Cameron, P.A., Dent, A.W., Staff, I.A., Suphioglu, C., 1999. Concentrations of major grass group 5 allergens in pollen grains and atmospheric particles: implications for hay fever and allergic asthma sufferers sensitized to grass pollen allergens. Clin. Exp. Allergy 29, 633–641.
Sercombe, J.K., Green, B.J., Tovey, E.R., 2006. Recovery of germinating fungal conidia from the nasal cavity after environmental exposure. Aerobiologia 22, 295–304.
Simmons, E.G., 2007. Alternaria. An Identification Manual. CBS Fungal Biodiversity Series, Utrecht, Netherlands.
Stennett, P.J., Beggs, P.J., 2004. Alternaria spores in the atmosphere of Sydney, Australia, and relationships with meteorological factors. Int. J. Biometeorol. 49, 98–105.

Stull, R.B., 1985. A fair-weather Cumulus clouds classification scheme for mixed layer studies. J. Clim. Appl. Meteorol. 24, 49–56.
Sweeney, M., Kroutil, L., Recht, M., Bush, R., 1985. Kinetics of allergen release from alternaria (ALT). J. Allergy Clin. Immunol. 75 (1), 118.
Taylor, P.E., Flagan, R.C., Valenta, R., Glovsky, M.M., 2002. Release of allergens as respirable aerosols: a link between grass pollen and asthma. J. Allergy Clin. Immunol. 109 (1), 51–56.
Troutt, C., Levetin, E., 2001. Correlation of spring spore concentrations and meteorological conditions in Tulsa, Oklahoma. Int. J. Biometeorol. 45, 64–74.
Twaroch, T., Arcalis, E., Sterflinger, K., Stoger, E., Swoboda, I., Valenta, R., 2012. Predominant localization of the major *Alternaria allergen* Alt a 1 in the cell wall of airborne spores. J. Allergy Clin. Immunol. 129 (4), 1148–1149.
Twaroch, T., Curin, M., Valenta, R., Swoboda, I., 2015. Mold allergens in respiratory allergy: from structure to therapy. Allergy, Asthma & Immunology Research 7 (7), 205–220.
Vloutoglou, I., Fitt, B.D.L., Lucas, J.A., 1996. Germination of *Alternaria linicola* conidia on linseed: effects of temperature, incubation time, leaf wetness and light regime. Plant Pathol. (Oxf.) 45 (3), 529–539.
Wei, T., Simko, V., 2017. R Package "Corrplot": *Visualization of a Correlation Matrix* (Version 0.84). Avaiable from. https://github.com/taiyun/corrplot.
Womiloju, T.O., Miller, J.D., Mayer, P.M., Brook, J.R., 2003. Methods to determine the biological composition of particulate matter collected from outdoor air. Atmos. Environ. 37 (31), 4335–4344.
Woo, Ch, An, Ch, Xu, S., Yi, S.-M., Yamamoto, N., 2018. Taxonomic diversity of fungi deposited from the atmosphere. ISME J. 12, 2051–2060.
Yamamoto, N., Bibby, K., Qian, J., Hospodsky, D., Rismani-Yazdi, H., Nazaroff, W.W., Peccia, J., 2012. Particle-size distributions and seasonal diversity of allergenic and pathogenic fungi in outdoor air. ISME J. 6, 1801–1811.
Yamamoto, N., Nazaroff, W.W., Peccia, J., 2014. Assessing the aerodynamic diameters of taxon-specific fungal bioaerosols by quantitative PCR and next-generation DNA sequencing. J. Aerosol Sci. 78, 1–10.